Troubleshooting

Problem	Potential Cause(s)	Recommended Solution
No FISH signals are detected in the microscope.	 Reflected light shutter closed / stop slider in light path. 	Open shutter / move stop slider out of the light path.
	 Fluorescent lamp is switched off. 	 Switch on fluorescent lamp.
	 Wrong fluorescence filter is in light path. 	 Move correct filter into light path.
	 Objective is out of position. 	 Swing objective into light path.
	 Phototube is in camera position. 	 Direct light path to eyepieces.
Hybridization signals become weak after a while.	 The immersion oil has penetrated between the glass slide and cover glass. 	 Replace coverslip and DAPI/antifade. Use 24 x 32 mm² coverslip even if only a small region is hybridized.
Diffuse signals.	Preparation is not adequately illuminated.	Check optical pathway of microscope. Adjust the UV light properly. Check the lifetime of the UV lamp.
	Focus plane cannot be adjusted properly.	 Use enough immersion oil. Do not mix different immersion oils. Use immersion oil suitable for fluorescence.
	Antifade layer is too thick for focusing.	 Do not use too much DAPI/antifade. 10 μl per slide (24 x 32 mm² coverslip) are sufficient.
Weak signals.	Chromosome slide preparation is too old.	 Slides should not be older than two weeks.
	 Denaturation of chromosomes is not adequate. 	 Aging, baking or further fixation may inhibit the hybridization and is not recommended. Increase denaturation temperature up to 80°C.
	 A multi bandpass filter is used for viewing. 	 Use a dedicated single bandpass filter.
Weak aqua or green signals or	 DAPI intensity is too high resulting in crosstalk to AQUA filter or GREEN filter. 	Use DAPI/antifade of low concentration.
high diffuse background in green color channel.	 pH value of washing solutions is too low. 	 Ensure that pH value is between 7.0 and 7.5 of solutions. Some green fluorophores are very sensible to pH below 7.
High unspecific background	 Remaining cytoplasmic proteins of the cells may impair the hybridization. 	Pretreat slides with Pepsin.

Customer Support

Please contact MetaSystems Probes GmbH (contact details see below) or our authorized distributor in your country. All trademarks are property of their respective owners.

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Symbols Used

Symbol	Description		
RUO	This symbol marks a product as for "Research Use Only".	⚠	All warnings are marked by warning triangle with exclamation mark. Depending on their character they are supplemented with the words ATTENTION or CAUTION.
w	Manufacturer	REF	Reference number
Y	No of tests	LOT	Lot number
R	Expiration date	Ł	Temperature limitation for storage. Lower and upper limits are indicated.
UDI	Unique Device Identification	*	Keep away from sunlight. Indicates a device that needs protection from light sources.
	Danger		

MetaSystems Probes GmbH 1. Industriestr. 7, 68804 Altussheim, Germany, Tel: +49 (0)6205 292760 **Danger** Paster Ha51, Ha73 Paster Ha51, Ha73 Paster Ha51, Ha73 Paster Paster Paster Ha73 Paster Ha51, Ha74 Paster Ha51



For Professional Use Only

Further information available at www.metasystems-probes.com

Product	Label	Order No.	Pack Size
XCE 10 green	green	D-0810-050-FI	50µl

The XCE 10 DNA probe contains green labeled repetitive sequences specific for the chromosomes 10 centromeric region. MetaSystems XCE probes are optimized for 1 hour hybridization.

Probe Diagram:



Chromosome 10

Revision: RUO-RevD220412-220407

Materials Provided

50µl of XCE 10 green, the probe is dissolved in hybridization solution (formamide, dextran sulfate, saline-sodium citrate) and ready to use.

Intended Use

XCE 10 green is intended for fluorescence in-situ hybridization (FISH) for the analysis of chromosomal aberrations on fixed cytogenetic specimen. This probe is intended for research use only. This product is not intended for diagnostic use

Safety Instructions

All probes produced by MetaSystems Probes are for professional use only and should be used by qualified and trained personnel only. To ensure safe operation and reproducible results please observe the safety notices and caution signs below.

	DANGER
Contains:	Formamide
Hazard statements:	H360FD May damage fertility. May damage the unborn child. H351 Suspected of causing cancer.
	H373 May cause damage to organs through prolonged or repeated exposure.
Precaution statements:	P201 Obtain special instructions before use.
	P260 Do not breathe dust/fume/gas/mist/vapors/spray.
	P280 Wear protective gloves/protective clothing. P308+ P313 IF exposed or concerned: Get medical advice/attention.
	P501 Dispose of contents/container to in accordance with local/regional/national/international regulations.
	Special labeling: Restricted to professional users.
\wedge	CAUTION: Hot water bath and hot plates!
Z • \	For denaturation and hybridization hot water baths and hot plates are used with temperatures of >37°C. Be
	careful not to get in direct contact with hot surfaces or liquids.
Δ	Wear gloves and a lab coat. In case of contact with skin, cool immediately with cold water.
<u> </u>	ATTENTION: Good Laboratory Practice! Use in accordance with the principles of good laboratory practice.
	Use in accordance with the philoples of good laboratory plactice.

Storage and Handling

Probes should be stored in the dark at -20°C (±5°C). Probe performance has been shown to be unaffected for up to 20 freeze-thaw cycles.

Shipping

Products produced by MetaSystems Probes are shipped at room temperature.

Equipment Necessary but not Supplied

 Water bath with accurate 	 Thermometer 	• FI
temperature control	 pH meter, calibrated 	(s
 Humidified chamber 37°C (±1°C) 	Timer	• İn
 Variable micro-pipettes with 	 Freezer -20°C (±5°C) 	• In
volumes ranging from 1 µl to 1 ml,	 Coplin jars (glass or plastic) 	m
calibrated	Microcentrifuge	• D
 Hotplate 75°C (±1°C), with a solid 	Gloves	R
plate and accurate temperature	 Forceps 	• C
control up to 80°C	DAPI/antifade	2
-		

- Fluorescence microscope with suitable filters
- (see below) Imaging System
- Imaging System
 Immersion oil, recommended by the microscope manufacturer (fluorescence grade)
- DAPI/antifade
- Rubber Cement
- Coverslips (glass):
- 22 x 22 mm² and 24 x 32 mm²

Fluorescence Microscope Recommendation

- Fluorescence Illumination: Metal halide fluorescence illumination systems or conventional 100 watt mercury lamp illuminators.
- Objectives: 10x/20x and 63x/100x suitable for epi-fluorescent illumination.
- Fluorescence Filters: For viewing/counting use an appropriate triple or quad bandpass filter set or appropriate single bandpass filter. For capturing images use suitable single bandpass filters for the respective fluorochromes. Please inquire.

Fluorochrome Specification

Label	Absorption max.	Emission max.	
Blue (aqua)	426 nm	480 nm	
Green	505 nm	530 nm	
Gold	525 nm	551 nm	
Orange	552 nm	576 nm	
Red	595 nm	615 nm	
Near Infrared	644 nm	669 nm	

Absorption and Emission depending on the label used in the product.

Sample Preparation

General Comments

- MetaSystems probes are designed for use on cytogenetic samples which are fixed in 3:1 methanol/acetic-acid and should be prepared according to the laboratory or institution guidelines.
- · Prepare specimen according to standard cytogenetic procedures.

Stability of Hybridized Slides

Hybridized FISH slides can be analyzed for at least six months if stored in the dark at -20°C (±5°C).

Additional Procedural Recommendations

- The use of a calibrated thermometer is strongly recommended for measuring temperatures of solutions, water baths, and incubators, as these temperatures are critical for optimum product performance.
- · Carefully check the temperature of preheated solutions.
- Carefully check the pH value of all solutions. It must be in the range of 7.0 7.5 at room temperature.
- The wash concentrations (stringency), pH and temperature are important, as low stringency can result in non-specific binding of the probe and too high stringency can result in a lack of signal.
- Before opening: Spin briefly to collect probe mix at the bottom of the tube.

FISH Protocol for DNA Probes

Slide Preparation

- 1. Spot cell sample onto cleaned microscope slide. Allow to air dry. If you are not using the slides the same day, store at -20°C (±5°C).
- 2. Apply 10 µl of probe mixture.
- 3. Cover with coverslip 22 x 22 mm².
- 4. Seal with rubber cement.

Denaturation

1. Denature sample and probe simultaneously by heating slide on a hotplate at 75°C (±1°C) for 2 min.

Hybridization

1. Incubate in a humidified chamber at 37°C (±1°C) for 1 hour to overnight.

Post-Hybridization Washes

Solutions Required

- 0.4X SSC (pH 7.0 7.5) at 72°C (±1°C)
- 2X SSC, 0.05% Tween-20 (pH 7.0) at room temperature

Procedure

- 1. Remove coverslip and all traces of glue carefully.
- 2. Wash slide in 0.4X SSC (pH 7.0) at 72°C (±1°C) for 2 min.
- 3. Drain slide and wash in 2X SSC, 0.05% Tween-20 (pH 7.0) at room temperature for 30 seconds
- 4. Rinse briefly in distilled water to avoid crystal formation and let air dry.

Counterstain

- Solutions required:
- DAPI/antifade

Procedure:

- 1. Apply 10 µl of the DAPI/antifade and overlay with a 24 x 32 mm² coverslip.
- 2. Allow penetration of DAPI/antifade for 10min
- 3. Proceed with microscopy and analysis.
- 4. Store slides at -20°C (±5°C). Hybridization signals are fine for at least six months.

Expected Results

XCE 10 green hybridized to a normal human metaphase spread the centromeric regions of the respective two homologue chromosomes show a distinct hybridization signal each. In normal interphase nuclei each centromeric region of the respective chromosome is represented by a distinct spot resulting in a total of 2 spots. Gain or loss of spots indicates numerical aberrations of the respective chromosome. Faint cross-hybridizations to the centromeric regions of other chromosomes may be observed. They can be reduced by increasing the stringency of the post-hybridization washings through higher temperature (74°C for 4 min) or lower salt concentration (0.25xSSC).