**Troubleshooting**

<table>
<thead>
<tr>
<th>Problem</th>
<th>Potential Cause(s)</th>
<th>Recommended Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td>No FISH signals are detected in the microscope.</td>
<td>• Reflected light shutter closed / stop slider in light path.</td>
<td>• Open shutter / move stop slider out of the light path.</td>
</tr>
<tr>
<td></td>
<td>• Fluorescent lamp is switched off.</td>
<td>• Switch on fluorescent lamp.</td>
</tr>
<tr>
<td></td>
<td>• Wrong fluorescence filter is in light path.</td>
<td>• Move correct filter into light path.</td>
</tr>
<tr>
<td></td>
<td>• Objective is out of position.</td>
<td>• Swing objective into light path.</td>
</tr>
<tr>
<td></td>
<td>• Phototube is in camera position.</td>
<td></td>
</tr>
<tr>
<td>Hybridization signals become weak after a while.</td>
<td>• Immersion oil soaked in-between slide and coverslip.</td>
<td>• Replace coverslip and DAPI/antifade. Use 24 x 32 mm² coverslip even if only a small region is hybridized.</td>
</tr>
<tr>
<td>Diffuse signals.</td>
<td>• Preparation is not adequately illuminated.</td>
<td>• Check optical pathway of microscope. Adjust the UV light properly. Check the lifetime of the UV lamp.</td>
</tr>
<tr>
<td></td>
<td>• Focus plane cannot be adjusted properly.</td>
<td>• Use enough immersion oil. Do not mix different immersion oils. Use immersion oil suitable for fluorescence.</td>
</tr>
<tr>
<td></td>
<td>• Antifade layer is too thick for focusing.</td>
<td>• Do not use too much DAPI/antifade. 10 µl per slide (24 x 32 mm² coverslip) are sufficient.</td>
</tr>
<tr>
<td>Weak signals.</td>
<td>• Chromosome slide preparation is too old.</td>
<td>• Slides should not be older than two weeks.</td>
</tr>
<tr>
<td></td>
<td>• Denaturation of chromosomes is not adequate.</td>
<td>• Increase denaturation temperature up to 80°C.</td>
</tr>
<tr>
<td></td>
<td>• A multi bandpass filter is used for viewing.</td>
<td>• Use a dedicated single bandpass filter.</td>
</tr>
<tr>
<td>Weak aqua or green signals or high diffuse background in green color channel.</td>
<td>• DAPI intensity is too high resulting in crosstalk to AQUA filter or GREEN filter.</td>
<td>• Use DAPI/antifade of low concentration.</td>
</tr>
<tr>
<td></td>
<td>• pH value of washing solutions is too low.</td>
<td>• Ensure that pH value is between 7.0 and 7.5 of solutions. Some green fluorophores are very sensible to pH below 7.</td>
</tr>
<tr>
<td>High unspecific background</td>
<td>• Remaining cytoplasmic proteins of the cells may impair the hybridization.</td>
<td>• Pretreat slides with Pepsin.</td>
</tr>
</tbody>
</table>

If the recommended measures do not solve the problem, or your problem is not listed, please contact MetaSystems Probes.

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**Customer Support**

Please contact MetaSystems Probes GmbH (contact details see below) or our authorized distributor in your country. MetaSystems Probes disclaims any proprietary interest in the marks and names of others.

**Symbols Used**

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>![Symbol]</td>
<td>This symbol marks a product as an &quot;In Vitro Diagnostic Medical Device&quot;.</td>
</tr>
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</tr>
<tr>
<td>![Symbol]</td>
<td>Reference number</td>
</tr>
<tr>
<td>![Symbol]</td>
<td>No of tests</td>
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<tr>
<td>![Symbol]</td>
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<tr>
<td>![Symbol]</td>
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Revision: RevB170222-180528v10.1

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**XCyting Locus-Specific Probes**

For Professional Use Only

Further information available at www.metasystems-probes.com

<table>
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<tr>
<th>Product</th>
<th>Label</th>
<th>Order No.</th>
<th>Pack Size</th>
</tr>
</thead>
<tbody>
<tr>
<td>XL t(7;12) MNX1/ETV6</td>
<td>orange/green</td>
<td>D-5101-100-OG</td>
<td>100µl</td>
</tr>
</tbody>
</table>

The XL t(7;12) MNX1/ETV6 is designed as a dual fusion probe. The green labeled probes flank the breakpoint at 12p13 (ETV6), and the orange labeled probes flank the breakpoint at 7q36 (MNX1 formerly HXB9).

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| Chromosome 7 | Chromosome 12 |
Materials Provided
100µl of XL t(7;12) MNX1/ETV6, the probe mix is dissolved in hybridization solution and ready to use.

Intended Use
DNA FISH probes are intended for fluorescence in-situ hybridization (FISH) for the analysis of chromosomal aberrations on fixed cells from human tissue suitable for cytogenetic investigation. Hybridized to metaphase and/or interphase nucleus FISH probes allow the analysis of chromosome structure or copy number variations to detect acquired genetic alterations according to the Global Medical Device Nomenclature (GMDN) CT929. FISH analysis is used as an adjunct test to other diagnostic investigations and not to be used as sole base for diagnosis or therapy decisions.

Safety Instructions
All probes produced by MetaSystems Probes are for professional use only and should be used by qualified and trained personnel only. In order to ensure safe operation and reproducible results please observe the safety notices and caution signs below.

CAUTION: Formamide is toxic and a potential teratogen!
Formamide is toxic and a teratogen. May cause harm to the unborn child. Do not breathe vapours; avoid skin contact! Wear gloves and a lab coat. In case of contact with skin or eyes, wash immediately with water.

CAUTION: Hot water bath and hot plates!
For denaturation and hybridization hot water baths and hot plates are used with temperatures of >37°C. Be careful not to get in direct contact with hot surfaces or liquids. Wear gloves and a lab coat. In case of contact with skin, cool immediately with cold water.

ATTENTION: Good Laboratory Practice!
Use in accordance with the principles of good laboratory practice.

ATTENTION: Waste Disposal!
All hazardous materials should be disposed of according to local/national regulation for hazardous waste disposal.

Storage and Handling
Probes should be stored in the dark at -20°C (±5°C). Probe performance has been shown to be unaffected for up to 20 freeze-thaw cycles.

Shipping
MetaSystems’ DNA probes are shipped at room temperature.

Equipment Necessary but not Supplied
- Water bath with accurate temperature control
- Variable micro-pipettes with volumes ranging from 1 µl to 1 ml, calibrated
- Thermometer
- pH meter, calibrated
- Timer
- Coplin jars (glass or plastic)
- Microcentrifuge
- Fluorescence microscope with suitable filters (see below)
- Hotplate 70°C (±1°C), with a solid plate and accurate temperature control up to 80°C
- Freezer -20°C (±5°C)
- Humidified chamber 37°C (±1°C)
- Forcips
- Gloves
- Immersion oil, recommended by the microscope manufacturer (fluorescence grade)
- Imaging System, e.g. Isis (MetaSystems)
- Coverslips (glass);
- 22 x 22 mm² and 24 x 32 mm²
- Rubber Cement
- DAPI/antifade
- Timer
- pH meter, calibrated
- Microcentrifuge
- Thermometer
- Variable micro-pipettes with volumes ranging from 1 µl to 1 ml, calibrated
- Coplin jars (glass or plastic)
- Hotplate 70°C (±1°C), with a solid plate and accurate temperature control up to 80°C
- Freezer -20°C (±5°C)
- Humidified chamber 37°C (±1°C)
- Forcips
- Gloves
- IMMERSION Oil
- Imaging System, e.g. Isis (MetaSystems)
- COverSliPs (glass);
- 22 x 22 mm² and 24 x 32 mm²
- RubBerk eMent
- DAPI/antifade
- Timer
- pH meter, calibrated
- Microcentrifuge
- Thermometer
- Water bath with accurate temperature control
- Variable micro-pipettes with volumes ranging from 1 µl to 1 ml, calibrated
- Immersion oil, recommended by the microscope manufacturer (fluorescence grade)
- Imaging System, e.g. Isis (MetaSystems)
- Coverslips (glass);
- 22 x 22 mm² and 24 x 32 mm²
- Rubber Cement
- DAPI/antifade
- Fluorescence microscope with suitable filters (see below)
- Hotplate 70°C (±1°C), with a solid plate and accurate temperature control up to 80°C
- Freezer -20°C (±5°C)
- Humidified chamber 37°C (±1°C)
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- Freezer -20°C (±5°C)
- Humidified chamber 37°C (±1°C)
- Forcips
- Gloves

Fluorescence Microscope Recommendation
- Fluorescence Illumination: Metal halide fluorescence illumination systems or conventional 100 watt mercury lamp illuminators
- Objects suitable for epi-fluorescent illumination.
- Fluorescence Filters: For viewing/counting use a MetaSystems triple or quad bandpass filter set or appropriate single bandpass filter.
- For capturing images use suitable single bandpass filters for the respective fluorochromes. Please inquire.

Fluorochrome Specification

<table>
<thead>
<tr>
<th>Label</th>
<th>Absorption max.</th>
<th>Emission max.</th>
</tr>
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<tbody>
<tr>
<td>Blue (aq)</td>
<td>420 nm</td>
<td>480 nm</td>
</tr>
<tr>
<td>Green</td>
<td>505 nm</td>
<td>530 nm</td>
</tr>
<tr>
<td>Orange</td>
<td>552 nm</td>
<td>576 nm</td>
</tr>
</tbody>
</table>

Sample Preparation

General Comments
- MetaSystems probes are designed for use on cytogenetic samples which are fixed in 3:1 methanol/acetic-acid and should be prepared according to the laboratory or institution guidelines.
- Prepare specimen according to standard cytogenetic procedures.

Stability of Hybridized Slides
- Hybridized FISH slides can be analyzed for at least six months if stored in the dark at -20°C (±5°C).

Additional Procedural Recommendations
- The use of a calibrated thermometer is strongly recommended for measuring temperatures of solutions, water baths, and incubators, as these temperatures are critical for optimum product performance.
- Carefully check the temperature of preheated solutions.
- Carefully check the pH value of all solutions. It must be in the range of 7.0 - 7.5 at room temperature.
- The wash concentrations (stringency), pH and temperature are important, as low stringency can result in non-specific binding of the probe and too high stringency can result in lack of signals.
- Before opening: Spin briefly to collect probe mix at the bottom of the tube.

FISH Protocol for MetaSystems’ DNA Probes

Slide Preparation
1. Spot cell sample onto cleaned microscope slide. Allow to air dry. If you are not using these slides the same day, store at -20°C (±5°C).
2. Apply 10 µl of probe mixture.
3. Cover with coverslip 22 x 22 mm².
4. Seal with rubber cement.

Denaturation
1. Denature sample and probe simultaneously by heating slide on a hotplate at 70°C (±1°C) for 2 min.

Hybridization
1. Incubate in a humidified chamber at 37°C (±1°C) overnight.

Post-Hybridization Washes

Solutions Required:
- 0.4 x SSC (pH 7.0 – 7.5) at 72°C (±1°C)
- 2 x SSC, 0.05% Tween-20 (pH 7.0) at room temperature

Procedure:
1. Remove coverslip and all traces of glue carefully.
2. Wash slide in 0.4 x SSC (pH 7.0) at 72°C (±1°C) for 2 min.
3. Drain slide and wash in 2 x SSC, 0.05% Tween-20 (pH 7.0) at room temperature for 30 seconds.
4. Rinse briefly in distilled water to avoid crystal formation and let air dry.

Counterstain

Solutions required:
- DAPI/antifade (e.g. MetaSystems DAPI/antifade, D-0902-500-DA)

Procedure:
1. Apply 10 µl of the DAPI/antifade and overlay with a 24 x 32 mm² coverslip.
2. Allow the penetration of DAPI/antifade for 1 min.
3. Proceed with microscoping and analysis.
4. Store slides at -20°C (±5°C). Hybridization signals are fine for at least six months.

Expected Results

Normal Cell:
- Two green (2G) and two orange (2O) signals.

Aberrant Cell (typical results):
- One green (1G), one orange (1O), and two green-orange fusion (2GO) (adjacent green and orange) signals.

Only the most frequent signal constellations are shown, other relevant signal patterns may be observed.